

CYANOGENIC GLYCOSIDES AS A POTENTIAL BIOREGULATOR

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Review



ABSTRACT

Natural substances which are considered to be a food that provides medical and health benefits are called bioregulator. Bioregulators can be used in medicine for preventing and treating migraine, hypertension, chronic inflammation, and other reaction source diseases. Amygdalin is considered for one of the most important bioregulator. It is a controversial nature cyanogenic glycoside abundant in the seeds of *Rosaceae* family. The family includes herbs, shrubs, and trees and most species are deciduous, but some are evergreen. In the past few years, has been a renewed interest about distribution of *Rosaceae* fruits because amygdalin has been used for many years in traditional and alternative medicine. Therefore, it is included in regulatory pathways and processes. Recent data indicate potential regulatory activity of amygdalin in signaling pathways of highly metastatic cells, suggesting that amygdalin might not only be an innovative tool to neutralize metastatic dissemination but also to complement mTOR-inhibitor based regimens.

Keywords: Amygdalin, cyanogenic glycosides, apoptosis, mTOR pathway

INTRODUCTION

In recent years considerable effort has been made to identify the metabolic factors linking nutrition with partial physiological processes (Monget and Martin, 1997; Kolesárová et al., 2011). Numerous data from various mammalian species have shown that natural substances may influence the physiological functions (Prunier and Quesnel, 2000) or on the other side tumorigenesis, carcinogenesis, angiogenesis (Subash et al., 2010; Aggarwal et al., 2009). The most widely plant constituents, which are biologically active and provide medical and health benefits are called bioregulators (Brower, 1998; Zeisel, 1999). The function of bioregulators is also important to examine from viewpoint of prevention of many reproductive alterations (Medved'ová et al., 2010). Neuroendocrine regulation manages reproductive system through axis hypothalamic, pituitary and gonads (Javorka et al., 2012). Sex hormones (female-estrogens, progesterone, male-androgens: testosterone) are steroids (fat soluble compounds) that control sexual maturity and reproduction. The endocrine glands ovaries, testes, or adrenal cortex regulate sexual development of an organism and affects the growth or function of the reproductive organs. The endocrine and extracellular signalling systems provide a means of communication between distant organs via the circulatory system, specific cell populations, neighbouring cell populations, and the external and internal environments (Chedrese, 2009). Fruits and vegetables contain many different natural components, some essential nutrients and also contain a variety of bioactive substances, which have other beneficial health effects (Kris-Etherton et al., 2002). In the past few years, there has been a renewed interest in evaluating the bioregulator content and distribution in patterns of fruits and vegetables (Flood et al., 2002; Ruiz et al., 2006).

Rosaceae family

Family *Rosaceae*, comprised of over 100 genera and 3000 species, is the third most economically important plant family in temperate regions (Dirlewanger et al., 2002). *Rosaceae* contain edible members such as almonds, apples, plums, peaches, pears, raspberries, sour cherries, sweet cherries, and strawberries. Other non-edible species with almost exclusively ornamental value include rose, hawthorn, potentilla, cotoneaster, and pyracantha. The products of this family are in high demand for their nutritional and esthetic values edible (Vavilov, 1951). *Rosaceae* fruits are also a major human dietary source of phytochemicals, such as flavonoids, cyanogenic glycosides, phytoestrogens (Mazur et al., 2000), and phenols that could potentially yield health and disease - fighting advantages. L-

Ascorbic acid, quercetin, kaempferol, myricetin, p-coumaric acid, gallic acid, and ellagic acid are well known antioxidants and/or cancer-inhibiting compounds that have been identified in these fruits (Macheix et al., 1991, 1998; Selmar, 1999). The family *Rosaceae* has found for own rich generic representation of an application in the prevention and treatment of many pathological conditions. In the past few years, has been a renewed interest about distribution of these fruits (Fakuta et al., 2003; Chang et al., 2005). *In vitro* and *in vivo* studies on animal models provide evidence that fruit and leaf extracts from many *Rosaceae* species inhibit some cancers or have strong antioxidant activities (Yau et al., 2002). Seeds of *Rosaceae* fruits contain a substantial amount of dietary protein (Nout et al., 1995) along with significant amounts of oil and fiber but this part of apricot also depending on the variety, contain the toxic cyanogenic glycoside - amygdalin (Gomes et al., 1998). Islamiyat et al. (2014) have developed and applied a high performance liquid chromatographic procedure for amygdalin quantification to investigate extraction efficiency and to determine levels in a range of commercially-available foods. Their results showed that seeds from *Rosaceae* species contained relatively high amounts (range 0.1–17.5 mg/g) of amygdalin compared to seeds from *non-Rosaceae* species (range 0.01–0.2 mg/g) (Yildirim et al., 2010). The apricot (*Prunus armeniaca* L.) is a member of the *Rosaceae* family. Apricot fruit, being a rich source of vitamins and minerals and is one of the most familiar crops worldwide. Their trees are not ubiquitous since they can only grow in certain regions where the environmental conditions are favourable (Baytop et al., 1999). The fresh apricot fruit contains carbohydrates, vitamins C and K, β-carotene, niacin, and thiamine. Organic acids, phenols, volatile compounds, esters, and terpenoids have also been isolated (Ruiz et al., 2006; Riu-Aumatell et al., 2005; Safer et al., 2006).

Bioactive substance - Amygdalin

Natural plant origin products like amygdalin are still a major part of traditional medicine (Hwang et al., 2008). Amygdalin (vitamin B17; previously called laetrile) is one of many nitrilosides, which are natural cyanide-containing substances abundant in the seeds of prunasin family and other *Rosaceae* plants (Chang et al., 2005; Pak et al., 1999). The distribution of the cyanogenic glycosides in the plant kingdom is relatively wide and they are present mainly in more than 2650 plant species (Franciscu and Pinotti, 2000; Haque and Bradbury, 2002). There are approximately 25 cyanogenic glycosides which have found in the edible parts of plants being: amygdalin (almonds); dhurrin (sorghum); linamarin (cassava, lima beans); lotaustralin (cassava, lima beans); prunasin (stone fruit); and taxiphyllin (bamboo shoots) (Gonzales and Sabatini,

1989). Cyanide is a toxic substance, mainly due to its affinity for the terminal cytochrome oxidase in the mitochondrial respiratory pathway (Brattsten et al., 1983). The lethal dose of cyanide for vertebrates lies in the range of 35–150 µmol/kg, if applied in a single dose. Much higher amounts of HCN can be tolerated if consumed or administered over a longer period (Davis and Nahrstedt, 1985). Biosynthesis and degradation of cyanogenic glycosides (CNGs) are well documented in many plants (Jones et al., 2000; Lechtenberg and Nahrstedt, 1999).

But the genetic control of cyanogenesis has no unique mechanism, the plants show variation in the amount of the produced HCN. The production of HCN depends on both the biosynthesis of CNGs and on the existence (or absence) of its degrading enzymes. The biosynthetic precursors of the CNGs are different L-amino acids, these are hydroxylated then the *N*-hydroxylamino acids are converted to aldoximes, these are turned into nitriles. The last ones are hydroxylated to α -hydroxynitriles and then they are glycosylated to CNGs. The generation of HCN from CNGs is a two steps process involving a deglycosylation and a cleavage of the molecule (regulated by β -glucosidase and α -hydroxynitrilase). The tissue level compartmentalisation of CNGs and their hydrolysing enzymes prevents large-scale hydrolysis in intact plant tissue. The actual level of CNGs is determined by various factors both developmental and ecological ones, which are reviewed too (Vetter, 2000).

Amygdalin is composed of two molecules of glucose, one of benzaldehyde, which induces an analgesic action, and one of hydrocyanic acid, which is an anti-neoplastic compound (Zhou et al., 2012).

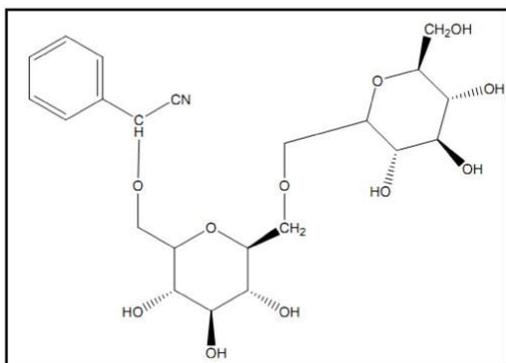


Figure 1 Chemical structure of amygdalin (Abdel-Rahman, 2011).

Action of endogenous plant enzymes can release hydrogen cyanide causing potential toxicity issues for animals and humans, including cell death by blocking cytochrome oxidase and the arrest of the ATP production (Bolarinwa et al., 2014). Amygdalin has been used to treat cancers and relieve pain (Ellison et al., 1978; Shim et al., 2000). Amygdalin was reported to selectively kill cancer cells at the tumor site without systemic toxicity and to effectively relieve pain in cancer patients (Zhou et al., 2012). The acute toxicity experiments of amygdalin have proved that the toxicity of oral administration route is far greater than the intravenous route (Adewusi and Oke, 1985; Park et al., 2013). The maximum tolerance dose of intravenous and intramuscular injection of amygdalin in mice, rabbits, dogs are 3g/kg, 0.075 g/kg orally respectively (Zhang and Jin, 1986; Rauws et al., 1982) and human intravenous injection are 5g (approximately 0.07 g/kg). Previous studies on amygdalin have focused on its purification, toxicity related to the release of cyanide, anti-tumor mechanism, and identification of its metabolites in plasma or herbs, and its pharmacological effect on cancers (Rauws et al., 1982, Song and Xu., 2014). Recent studies examined the effects of natural compound amygdalin on female reproductive system concentrated on secretory activity of porcine ovarian granulosa cells (GC) *in vitro* (Halenár et al., 2013a). Halenár et al. (2015) have investigated the release of steroid hormone progesterone by GC from cyclic and non-cyclic porcine ovaries. The progesterone release was not significantly ($P > 0.05$) affected by the amygdalin treatment at all experimental doses (1, 10, 100, 1000 and 10000 mg/ml) compared to the control group without addition. However, a significant stimulation ($P < 0.05$) of the 17- β -estradiol release after amygdalin addition at the highest dose (10000 mg/ml) was observed. Other experimental doses of amygdalin (1, 10, 100 and 1000 mg/ml) did not cause differences in the 17- β -estradiol secretion. Kolesár et al., (2015) in their review have described the characteristic, metabolism and possible effects of amygdalin on reproductive processes. Previous studies described the effects of natural compound amygdalin on female and male reproductive systems focused on process of steroidogenesis (Halenár et al., 2013a, 2015), spermatozoa motility and morphological abnormalities of bull spermatozoa (Tanyildizy and Bozkurt, 2004). Amygdalin significantly inhibited sperm hyaluronidase activity. The inhibition of hyaluronidase activity can cause a drop in the fertilization ability of bull spermatozoa due to the prevention of acrosomal reaction. However, amygdalin did not produce any morphological abnormality in bull spermatozoa. The

inhibition of sperm hyaluronidase activity and spermatozoa motility showed that these compounds have deleterious effects on bull sperm *in vitro* (Tanyildizy and Bozkurt, 2004).

Amygdalin is one of main pharmacological components of crude ingredients of Keishi-bukuryo-gan, Japanese herbal medicine (Yasui et al., 2003). It has been used for induction of ovulation in women suffering from infertility (Igarashi, 1988). Keishi-bukuryo-gan and its crude ingredients affected steroidogenesis in pre-ovulatory follicles (Usuki, 1987, 1990, 1991) and the corpus luteum (Usuki, 1986, 1988) in the rat ovary *in vivo* and *in vitro*.

The characterization and role of mTOR

Mammalian target of rapamycin (mTOR) is a serine/threonine kinase, which belongs to phosphatidylinositol-3 kinase (PI3K) related kinases (PIKKs) family. It regulates cellular metabolism, growth, proliferation, and therefore is a target for the development of a number of mTOR inhibitors (Pópulo et al., 2012). Akt functions as a component of the PI3K cell survival pathway. In cancer, Akt activity is frequently elevated due to multiple mechanisms, including loss of function of the PTEN tumor-suppressor gene and mutations of the PIK3CA gene. Akt functions as a component of the PI3K cell survival pathway (Manning et al., 2005). Akt acts as a survival kinase in many cancers (Cheng et al., 2005). The PI3K pathway is implicated in cell survival and cell growth, and can be activated by growth factors binding to cell-surface receptors. It is an intricate signaling cascade that is among the most frequently activated pathways in different types of cancer. PI3K is the subject of extensive research (Knight et al., 2007).

Qian et al. (2015) submitted a study where Western blot results showed that amygdalin had no significant impact on Akt and Rictor expression. Rictor as a subunit of mTOR plays an important role in the Akt-mTOR signaling pathway; its phosphorylation level is positively regulated by Akt (Chen et al., 2010). But the determination of Akt and rictor phosphorylation level showed that amygdalin significantly reduced the phosphorylation level of these two proteins in highly metastatic cells, suggesting that amygdalin was able to regulate the activity of Akt and rictor signaling pathways (Qian et al., 2015). The PI3K pathway is a key signal transduction system that links oncogenes and multiple receptor classes to many essential cellular functions, and is perhaps the most commonly activated signalling pathway in human cancer. This pathway therefore presents both an opportunity and a challenge for cancer therapy. Even as inhibitors that target PI3K isoforms and other major nodes in the pathway, including Akt and mTOR, reach clinical trials, major issues remain. Liu et al., (2009) describe the progress made in understanding of the PI3K pathway and discuss the potential of and challenges for the development of natural therapeutic agents that target this pathway in cancer (Liu et al., 2009). Thus, amygdalin might not only be an innovative tool to neutralize metastatic dissemination but also to complement mTOR-inhibitor based regimens (Gupta et al., 2013).

SUMMARY, CONCLUSIONS, AND FUTURE PERSPECTIVES

This review described possible effects of natural bioregulators on various types of animal cells. In recent years, increasing attention has been paid to natural substances and their impact on specific pathways in the cell. Amygdalin, as natural product shows lot of evidences. This natural compound is known for its anticancer, anti-inflammatory activity and other medicinal benefits, but on the other side represents one of the most controversial substance. Although amygdalin itself is non-toxic but its production HCN splitted by some endogenous plant enzymes is toxic substance for animals including humans. These agents that inhibit the downstream protein kinase mTOR as well as agents that inhibit multiple kinases, including components of the PI3K-Akt pathway are under clinical evaluation. There are still only a few studies which could suggest the possible involvement of amygdalin in mTOR pathway and thus influence animal reproductive system. Therefore other *in vitro* and *in vivo* experiments are necessary.

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REFERENCES

- Abdel-Rahman M.K. (2011). Can apricot kernels fatty acids delay the atrophied hepatocytes from progression to fibrosis in dimethylnitrosamine (DMN)-induced liver injury in rats? *Lipids in Health and Disease*. 10(1), 114.
- Adewusi, S. R., Oke, O.L. 1985. On The Metabolism Of Amygdalin. 2. The Distribution Of Beta-Glucosidase Activity And Orally Administered Amygdalin In Rats. *Canadian Journal of Physiology and Pharmacology*, .. 63, 1084-1087. <http://dx.doi.org/10.1139/y85-178>
- Aggarvall, B. B., Van Kuiken, M. E., Iyer, L. H. - Sung, B. (2009). Molecular Targets of Nutraceuticals Derived from Dietary Spices: Potential Role in Suppression of Inflammation and Tumorigenesis. *Experimental Biology and Medicine*, 234,8, 825-849. <http://dx.doi.org/10.3181/0902-mr-78>

- Baytop T. *Türkiyede bitkilerle tedavi*. Istanbul: Istanbul Eczacılık Fakültesi Yayınları; 1999. http://dx.doi.org/10.1501/ilhfak_0000001385
- Bolarinwa, I. F., Orfila, C., & Morgan, M. R. (2014). Amygdalin content of seeds, kernels and food products commercially-available in the UK. *Food chemistry*, 152, 133-139. <http://dx.doi.org/10.1016/j.foodchem.2013.11.002>
- Brattsten, L.B., Samuelian, J.H., Long, K.Y., Kincaid, S.A., Evans, C.K., 1983. Cyanide as a feeding stimulant for the southern armyworm, *Spodoptera eridania*. *Ecol. Entomology* 8, 125–132. <http://dx.doi.org/10.1111/j.1365-2311.1983.tb00490.x>
- Brower, V. (1998). Nutraceuticals: Poisoned for a healthy slice of the healthcare market? *Nature Biotechnology*, 16(8), 728-731. <http://dx.doi.org/10.1038/nbt0898-728>
- Calderaro J, Rebouissou S, de Koning L, Masmoudi A, He'rault A. (2014) PI3K/AKT pathway activation in bladder carcinogenesis. *International Journal of Cancer* 134 (8) 1767–2018. <http://dx.doi.org/10.1002/ijc.28518>
- Cantley LC. (2002). *Science*, 296, 1655–1657. <http://dx.doi.org/10.3390/ijms13021886>
- Chedrese J P. Reproductive endocrinology: a molecular approach., Springer., 2009, p. 382. <http://dx.doi.org/10.1007/978-0-387-88186-7>
- Davis, R.H., Nahrstedt, A., 1985. Cyanogenesis in insects. In: Kerkut, G.A., Gilbert, L.I. (Eds.), *Comprehensive Insect Physiology, Biochemistry and Pharmacology*. Pergamon Press, Oxford, pp. 635– 654. <http://dx.doi.org/10.1016/b978-0-08-030812-8.50020-4>
- Dirlewanger, E., Cosson, P., Tavaud, M., Aranzana, M.J., Poizat, C., Zanetto, A., Arús, P., Laigret, F., (2002) Development of microsatellite markers in peach [*Prunus persica* (L.) Batsch] and their use in genetic diversity analysis in peach and sweet cherry (*Prunus avium* L.). *Theoretical Applied Genetics*.105,127-138. <http://dx.doi.org/10.1007/s00122-002-0867-7>
- Ellison N. M., Byar D. P., Newell G. R. (1978). *Journal of Medicine*. 299, 549-552. <http://dx.doi.org/10.1056/nejm197809072991013>
- Eric E. Conn. (1969). Cyanogenic glycosid. *Journal of the Science of Food and Agriculture.*, 17 (3), 519-526 <http://dx.doi.org/10.1021/jf60163a014>
- Flood, A., Velie, E.M., Chatterjee, N., Subar, A.F., Thompson, F.E., Lacey, J.V. Jr. (2002). Fruit and vegetable intakes and the risk of colorectal cancer in the Breast Cancer Detection Demonstration Project follow-up cohort. *American Journal of Clinical Nutrition*, 75, 936-943. <http://dx.doi.org/10.1093/ajcn/dyg052>
- Fukuda T., Ito H., Mukainaka T., Tokuda H., Nishino H., Yoshida T. (2003). *Biological and Pharmaceutical Bulletin*. 26, 271-273. <http://dx.doi.org/10.1248/bpb.26.271>
- Franciscu, I. A., Pinotti, M. H. P. Cyanogenic glycosides in plants. *Braz. Arch. Biol. Technol.*, 2000, 43, 487–492. <http://dx.doi.org/10.1590/s1516-89132000000500007>
- Gomez, E., Burgos, L., Soriano C., Marin, J. (1998). Amygdalin content in the seeds of several apricot cultivars. *Journal of the Science of Food and Agriculture* 77, 184-186. [http://dx.doi.org/10.1002/\(sici\)1097-0010\(199806\)77:2<184::aid-jsfa22>3.0.co;2-h](http://dx.doi.org/10.1002/(sici)1097-0010(199806)77:2<184::aid-jsfa22>3.0.co;2-h)
- Gonzales, I. and Sabatini, S. (1989). Cyanide poisoning: pathophysiology and current approaches to therapy. *The International Journal of Artificial Organs*, 12(6): 347-355. <http://dx.doi.org/10.1111/aor.2000.24.issue-12>
- Gupta, S., Hau, A. M., Beach, J. R., Harwalker, J., Mantuano, E., Gonias, S. L., ... & Hansel, D. E. (2013). Mammalian target of rapamycin complex 2 (mTORC2) is a critical determinant of bladder cancer invasion. *Plos one* .8 (11), 3-9 <http://dx.doi.org/10.1371/journal.pone.0081081>
- Halenár, M., Medved'ová, M., Maruniaková, N., Packová, D., Kolesárová, A., (2013), Amygdalin and its effects on animal cells. *Journal of Microbiology and Biotechnology.*, 2 (1), 1414-1423 <http://dx.doi.org/10.15414/jmbfs.2015.4.special2.19-22>
- Halenár, M., Medved'ová, M., Maruniaková, N., Packová, D., Kolesárová, A., (2014). Dose-response of porcine ovarian granulosa cells to amygdalin treatment combined with deoxynivalenol. *Journal of Microbiology and Biotechnology.*, 4(2), 19-22. <http://dx.doi.org/10.15414/jmbfs.2015.4.special2.19-22>
- Halenár, M., Medved'ová, M., Maruniaková, N., Kolesárová, A., (2015), Assessment of a potential preventive ability of amygdalin in mycotoxin-induced ovarian toxicity. *Journal of Environmental Science and Health.*, 50, 411–416. <http://dx.doi.org/10.1080/03601234.2015.1011956>
- Haque, M. R., Bradbury, J. H. Total cyanide determination of plants and foods using the picrate and acid hydrolysis methods. *Food Chem.*, 2002, 77, 107–114. [http://dx.doi.org/10.1016/s0308-8146\(01\)00313-2](http://dx.doi.org/10.1016/s0308-8146(01)00313-2)
- Hwang, H., Lee, H., Kim, C., Shim, I., Hahm, D., (2008). Inhibitory effect of amygdalin on lipopolysaccharide-inducible TNF-alpha and IL-1beta mRNA expression and carrageenan-induced rat arthritis. *Journal of Microbiology and Biotechnology.*, 18(10), 1641-1647. <http://dx.doi.org/10.1248/bpb.31.1559>
- Chang H. K., Yang H. Y., Lee T. H., Shin M. C., Lee M. H., Shin M. S., Kim C. J., Kim O. J., Hong S. P., Cho S. (2005). *Biological and Pharmaceutical Bulletin.*, 28, 449-454. <http://dx.doi.org/10.1248/bpb.28.449>
- Chen, C. C., Jeon, S. M., Bhaskar, P. T., Nogueira, V., Sundararajan, D., Tonic, I., ... & Hay, N. (2010). FoxOs inhibit mTORC1 and activate Akt by inducing the expression of Sestrin3 and Rictor. *Developmental cell*, 18(4), 592-604. <http://dx.doi.org/10.1016/j.devcel.2010.03.008>
- Cheng JQ, Lindsley CW, Cheng GZ, Yang H, Nicosia SV.. (2005). The Akt/PKB pathway: molecular target for cancer drug discovery. *Oncogene*. 24,7482-7492 <http://dx.doi.org/10.1038/sj.onc.1209088>
- Igarashi, M. (1988). Kampo medicine in endocrinology. In *Recent Advances in the Pharmacology of Kampo (Japanese herbal) medicines*, E. Hosoya, Y. Yamamura (eds). Tokyo, *Excepta Medica*, 1988, 157-160. <http://dx.doi.org/10.1055/s-2006-960085>
- Javorka J., Béder I., Béderová A., Buc M., Čalkovská A., Donič V., Hájek J., Zeman M., (2012) *Lekárska fyziológia*. Osveta, Martin, p. 453.
- Jones, P.R., Andersen, M.D., Nielsen, J.S., Høj, P.B., Møller, B.L., 2000. The biosynthesis, degradation, transport and possible function of cyanogenic glucosides. In: Romero, J.T., Ibrahim, R., Varin, L., De Luca, V. (Eds.), *Evolution of Metabolic Pathways*. Elsevier Science, New York, pp. 191–247. [http://dx.doi.org/10.1016/s0079-9920\(00\)80008-8](http://dx.doi.org/10.1016/s0079-9920(00)80008-8)
- Kolesár, E., Halenár, M., Kolesárová A., Massányi P., (2015). Natural plant toxicant – cyanogenic glycoside amygdalin: characteristic, metabolism and the effect on animal reproduction. *Journal of microbiology, biotechnology and food sciences*. 4(2),49-50. <http://dx.doi.org/10.15414/jmbfs.2015.4.special2.49-50>
- Kolesárová, A., Capcarová, M., Baková, Z., Gálik, B., Juráček, M., Šimko, M., Sirotkin, A.V. (2011). The effect of bee pollen on secretion activity, markers of proliferation and apoptosis of porcine ovarian granulosa cells in vitro. *Journal of Environmental Science and Health, Part B*, 46(3) 207-212. <http://dx.doi.org/10.1080/03601234.2011.540202>
- Knight ZA, Shokat KM. (2007). Chemically targeting the PI3K family. *Biochem Soc Trans*, 35, 245-249.
- Kris-Etherton, P.M., Hecker, K.D., Bonanome, A., Coval, S.M., Binkoski, A.E., Hilpert, K.F., et al.(2002). Bioactive compounds in foods: their role in the prevention of cardiovascular disease and cancer. *American Journal Medicine*, 113, 71S-88. [http://dx.doi.org/10.1016/s0002-9343\(01\)00995-0](http://dx.doi.org/10.1016/s0002-9343(01)00995-0)
- Lamming, D. W., Mihaylova, M. M., Katajisto, P., Baar, E. L., Yilmaz, O. H., Hutchins, A., ... & Sabatini, D. M. (2014). Depletion of Rictor, an essential protein component of mTORC2, decreases male lifespan. *Aging cell*, 13(5), 911-917. <http://dx.doi.org/10.1111/accel.12256>
- Lechtenberg, M., Nahrstedt, A., 1999. Cyanogenic glucosides. In: Ikan, R. (Ed.), *Naturally Occurring Glycosides*. John Wiley & Sons, New York, pp. 147–191. [http://dx.doi.org/10.1002/\(sici\)1099-1026\(199911/12\)14:6<426::aid-fft846>3.0.co;2-9](http://dx.doi.org/10.1002/(sici)1099-1026(199911/12)14:6<426::aid-fft846>3.0.co;2-9)
- Liu P., Cheng H., Roberts T.M, Zhao J.J., (2009). Targeting the phosphoinositide 3-kinase pathway in cancer. *Nature Reviews* 8, 627-644. <http://dx.doi.org/10.1038/nrd2926>
- Manning BD, Cantley LC. (2007). AKT/PKB signaling: navigating downstream. *Cell*. 129, 1261-1274.
- Macheix, J.J., Sapis, J., Fleuriet, A., (1991). Phenolic compounds and polyphenoloxidase in relation to browning in grapes and wines. *Critical Reviews in Food Science and Nutrition* 30, 441-486. <http://dx.doi.org/10.1080/10408399109527552>
- Mazur, W., Uehara, M., Wahala, K., Adlercreutz, H., (2000). Phyto-oestrogen content of berries, and plasma concentrations and urinary excretion of enterolactone after asingle strawberry-meal in human subjects. *British Journal of Nutrition*. 83, 381-387. <http://dx.doi.org/10.1079/bjn19980007>
- Medved'ová M., Kolesárová A, Capcarová M., Sirotkin A., Kováčik J., (2010), The release of progesterone by ovarian granulosa cells following cobalt experimental administration *Potravinárstvo*, 4, p. 33-336. <http://dx.doi.org/10.1080/10934521003708968>
- Meyers, S.A., Rosenberger, A. E. 1999. A plasma membrane-associated hyaluronidase is localized to the posterior acrosomal region of stallion sperm and is associated with spermatozoa function. *Biology of Reproduction.*, 61(2), 444-451 <http://dx.doi.org/10.1095/biolreprod61.2.444>
- Moertel, C. G., Ames, M. M., Kovach, J. S., Moyer, T. P., Rubin, J. R., & Tinker, J. H. (1981). A pharmacologic and toxicological study of amygdalin. *Jama*, 245(6), 591-594. <http://dx.doi.org/10.1001/jama.245.6.591>
- Monget, P., Martin, G. B. (1997). Involvement of insuline-like growth factors in the interaction between nutrition and reproduction in female mammals. *Human Reproduction Volume*, 12 (1), 33-52. http://dx.doi.org/10.1093/humrep/12.suppl_1.33
- Pak J. U., Moon S. J., Moon K., Won J. H. (1999). *Journal of Korean Oriental Oncology*. 5, 137-150. <http://dx.doi.org/10.3346/jkms.1999.14.5.546>
- Nout MJ, Tuncel G, Brimer L. Microbial degradation of amygdalin of bitter apricot seeds (*Prunus armeniaca*). *Int J Food Microbiol*. 1995; 24: 407-412. [http://dx.doi.org/10.1016/0168-1605\(94\)00115-m](http://dx.doi.org/10.1016/0168-1605(94)00115-m)
- Pak JU., Moon SJ., Moon K., Won JH.(1999) Effects of Semen Armenicae and Radix Trichosanthis on the iNOS expression and superoxide formation in the RAW264.7 cells. *Journal of Korean Oriental Oncology*. 5,137-150. <http://dx.doi.org/10.4093/jkda.2006.30.5.336>
- Park, H. J., Yoon, S. H., Han, L. S., Zheng, L. T., Jung, K. H., Uhm, Y. K., et al. (2005). Amygdalin inhibits genes related to cell cycle in SNU-C4 human colon cancer cells. *World Journal of Gastroenterology.*, 11, 5156-5161.

- Pópulo, H., Lopes, J. M., & Soares, P. (2012). The mTOR signalling pathway in human cancer. *International journal of molecular sciences*, 13(2), 1886-1918. <http://dx.doi.org/10.3390/ijms13021886>
- Poulton J. E. (1990). Cyanogenesis in plants. *Review Plant Physiology*, 94, 401-405. <http://dx.doi.org/10.1104/pp.94.2.401>
- Rauws, A.G., Olling, M., Timmerman, A. (1982) The pharmacokinetics of amygdalin. *Archive of toxicology*, 49, 1982, 311-312. <http://dx.doi.org/10.1007/bf00347879>
- Prunier, A., Quesnel, H. (2000). Influence of the nutritional status on ovarian development in female pigs. *Animal Reproduction Science*, 60(61), 185-197. [http://dx.doi.org/10.1016/s0378-4320\(00\)00093-2](http://dx.doi.org/10.1016/s0378-4320(00)00093-2)
- Qian, L., Xie, B., Wang, Y., & Qian, J. (2015). Amygdalin-mediated inhibition of non-small cell lung cancer cell invasion in vitro. *International journal of clinical and experimental pathology*, 8(5), 5363. <http://dx.doi.org/10.1016/j.ajpath.2012.11.019>
- Ruiz, D., Egea, J., Gil, M.L., Tomas-Barberan, F.A., (2006). Phytonutrient content in new apricot (*Prunus armeniaca* L.) varieties. *Acta Horticulturae* 717, 363-368. <http://dx.doi.org/10.17660/actahortic.2006.717.73>
- Selmar, D., (1999). Cyanide in Foods. *Phytochemicals in human health protection, nutrition, and plant defense* (Vol.33).USA: Springer. http://dx.doi.org/10.1007/978-1-4615-4689-4_14
- Shim B. S., Park J. K., Choi S. H. (2000). *Journal of Korean Oriental Oncology*, 6, 19-28. <http://dx.doi.org/10.4070/kcj.2000.30.2.227>
- Shragg, T.A., Albertson, T.E., Fisher, C.J., Jr. Cyanide poisoning after bitter almond ingestion. (1982) *Western Journal of Medicine* .136, 65-69. <http://dx.doi.org/10.4103/1755-6783.140262>
- Subash, C., Gupta, J., Hye, K., Sahdeo, P., Bharat, B. (2010). Regulation of survival, proliferation; invasion, angiogenesis, and metastasis of tumor cells through modulation of inflammatory pathways by nutraceuticals. *Cancer Metastasis Reviews*, 29(3), 405-434. <http://dx.doi.org/10.1007/s10555-010-9235-2>
- Suchard, J. R., Wallace, K. L., Gerkin, R. D. (1998). Acute cyanide toxicity caused by apricot kernel ingestion. *Annals of Emergency Medicine*. 32, 742-744. [http://dx.doi.org/10.1016/s0196-0644\(98\)70077-0](http://dx.doi.org/10.1016/s0196-0644(98)70077-0)
- Song Z., Xu X. (2014) Advanced research on anti-tumor effects of amygdalin, 10(5), 3-7. <http://dx.doi.org/10.4103/0973-1482.139743>
- Tanyildizi, S., Bozkurt, T. (2004). In Vitro Effects of Linamarin, Amygdalin and Gossypol Acetic Acid on Hyaluronidase Activity, Sperm Motility and Morphological Abnormality in Bull Sperm. *Turkish Journal of Veterinary and Animal Sciences*, vol. 28, 2004, p. 819-824.
- Usuki, S. 1986. Effects of Chinese herbal medicines on progesterone secretion by corpus luteum. In *Japanese Journal of Fertility and Sterility*, vol. 31, 1986, p. 482-486. <http://dx.doi.org/10.1142/s0192415x86000260>
- Usuki, S. 1987. Effects of Hachimijiogan, Tokishakuyakusan and Keishibukuryogan on estrogen and progesterone secretion in ovarian follicles. In *Japanese Journal of Fertility and Sterility*, vol. 32, 1987, p. 276-283. <http://dx.doi.org/10.1142/s0192415x8700014x>
- Usuki, S. 1988. Effects of Hachimijiogan, Tokishakuyakusan and Keishibukuryogan on progesterone and 17 α -hydroxyprogesterone secretion by rat corpora lutea in vivo. In *Japanese Journal of Fertility and Sterility*, vol. 33, 1988, p. 60-66. <http://dx.doi.org/10.1142/s0192415x88000078>
- Usuki, S. 1990. Effects of Tokishakuyakusan and Keishibukuryogan on steroidogenesis by rat preovulatory follicles in vivo. In *American Journal of Chinese Medicine*, vol. 18, 1990, p. 149-156. <http://dx.doi.org/10.1142/s0192415x90000198>
- Usuki, S. (1991). Effects of Hachimijiogan, Tokishakuyakusan and Keishibukuryogan, Ninjinto and Unkeito on estrogen and progesterone secretion in preovulatory follicles incubated in vitro. *American Journal of Chinese Medicine*, 19, 1991, 65-71. <http://dx.doi.org/10.1142/s0192415x91000107>
- Vavilov, N. I. (1951). The origin, variation, immunity and breeding of cultivated plants. *Soil Science*, 72(6), 482. <http://dx.doi.org/10.1097/00010694-195112000-00018>
- Vetter, J. 2000 Plant cyanogenic glycosides, *Review Toxicol*, 38, p. 11-36. [http://dx.doi.org/10.1016/s0041-0101\(99\)00128-2](http://dx.doi.org/10.1016/s0041-0101(99)00128-2)
- Yan, Z., Denneboom, C., Hattendorf, A., Dolstra, O., Debener T, Stam P, Visser PB (2005) Construction of an integrated map of rose with AFLP, SSR, PK, RGA, RFLP, SCAR and morphological markers. *Theoretical Applied Genetics* 110, 766-777. <http://dx.doi.org/10.1007/s00122-004-1903-6>
- Yasui, T., Matsuzaki, T., Ushigoe, K., Kuwahara, A., Maegawa, M., Furumoto, H., AONO, T., Irahara, M. (2003). Stimulatory effect of the herbal medicine Keishi-bukuryo-gan on a cytokine-induced neutrophil chemoattractant, in rat ovarian cell culture. *American Journal of Reproductive Immunology*, 50, 2003, 90-97. <http://dx.doi.org/10.1034/j.1600-0897.2003.00055.x>
- Yau, M.H., Che, C.T., Liang, S.M., Kong, Y.C., Fong, W.P. (2002). An aqueous extract of *Rubus chingii* fruits protects primary rat hepatocytes against tert-butyl hydroperoxide induced oxidative stress. *Life Science* 72, 329-338. [http://dx.doi.org/10.1016/s0024-3205\(02\)02239-7](http://dx.doi.org/10.1016/s0024-3205(02)02239-7)
- Yildirim AN, San B, Koyuncu F, Yildirim F (2010). Variability of phenolics, α -tocopherol and amygdalin contents of selected almond (*Prunus amygdalus* Batsch.) genotypes. *Journal of Food Agriculture and Environment*, 8(1): 76-79. <http://dx.doi.org/10.1002/jsfa.2740350616>
- Zeisel, S. H. (1999). Regulation of nutraceuticals. *Science*, 285, 1853-1855. <http://dx.doi.org/10.1126/science.285.5435.1853>
- Zhang, G. M., Jin, B. Q. (1986). Pharmacokinetics of amygdalin in rabbits. *Zhongguo Yao Li Xue Bao*, 7, 460-462. <http://dx.doi.org/10.1086/591376>
- Zhou, C., Qian, L., Ma, H., Yu, X., Zhang, Y., QU, W., Zhang, X., Xia, W. (2012). Enhancement of amygdalin activated with β -D-glucosidase on HepG2 cells proliferation and apoptosis. *Carbohydrate Polymers*, 90., 516-523. <http://dx.doi.org/10.1016/j.carbpol.2012.05.073>