



## RELATIONSHIP BETWEEN LEVEL OF COPPER IN BOVINE SEMINAL PLASMA AND SPERMATOZOA MOTILITY

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### ABSTRACT

The aim of this study was to evaluate relationship between copper (Cu) concentration of bovine seminal plasma and spermatozoa motility. Semen samples were collected from 13 breeding bulls. The motility analysis was carried out using the Computer Assisted Sperm Analysis (CASA) system. The mean value for the percentage of motile spermatozoa (MOT) was  $92.46 \pm 3.99\%$  and the progressive motility of the spermatozoa (PROG) as  $90.23 \pm 4.02\%$ . The seminal plasma Cu concentrations were analyzed by UV/VIS spectrophotometry. The total Cu concentration of the seminal plasma was  $4.28 \pm 1.47 \mu\text{M/L}$ . The correlation analysis revealed a strong negative correlation between MOT and seminal plasma Cu concentration ( $r_p = -0.781$ ;  $P < 0.01$ ) as well as between PROG and Cu content in the seminal plasma ( $r_p = -0.726$ ;  $P < 0.01$ ). The data obtained from this study clearly indicated that concentration of copper in seminal plasma negatively affects the spermatozoa motility parameters and subsequently might cause reproductive alteration in male sexual functions.

**Keywords:** copper, bovine spermatozoa, semen analysis, seminal plasma, motility parameters

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## INTRODUCTION

The natural environmental factors and differentiated anthropogenic pollutants, as well as many other sources strongly influence the reproductive material located in the semen, both in animals and humans (Fergusson, 1990; Marzec-Wroblewska et al., 2012). Mammalian semen is known to contain a big variety of chemical elements (Marzec-Wroblewska et al., 2012), whose influence on spermatozoa viability has been extensively studied in animals as well as in humans (Kanwal et al., 2000; Massányi et al., 2003a,b; Eghbali et al., 2008; Atig et al., 2012). Some of them are crucial for a proper sperm cell function (e. g. sodium, magnesium, calcium, potassium), others are required in relatively narrow limits (zinc, manganese, copper, iron, selenium) (Massányi et al., 2003a,b; 2004; Tvrdá et al., 2012).

The possible influence of metals ions, and especially copper (Cu), on the male infertility is a matter of great interest. Copper is an important trace and essential element for the all organisms (Craig et al., 2009), because has a great positive role in physiological and regulatory processes (Dobrzanski et al., 1996). Moreover, it is a component of a number of metalloenzymes and metalloproteins (Agarwal et al., 1990), which are involved in energy and antioxidant metabolism (Haliwell and Gutteridge, 2000; Aydemir et al., 2006).

Copper is a normal constituent of semen bound to the tail midpiece of spermatozoa (Manu, 1974) and present in seminal plasma, ampullar and seminal vesicular fluids and also released by other structures of the reproductive system (i.e. epididymis) (Valsa et al., 1994). Copper deficiency affects the development of sperm cells (Van Niekerk and Van Niekerk, 1989; Leonhard-Marek, 2000). On the other hand, the high doses of copper ions (Cu<sup>2+</sup>) have a toxic effect on the epididymis (Xu et al., 1985), testes, scrotum of mammals (Skandhan, 1992; Eidi et al., 2010), which may ultimately lead to a reduced fertility (Pesch et al., 2006). Increased levels of metal ions in semen (Umeyama et al., 1986) or seminal plasma (Stanwell-Smith et al., 1983) appear to be significantly and positively correlated with male infertility. Meeker et al. (2008) found evidence of an inverse association between high Cu levels and semen quality, which is consistent with a number of animal and human studies (Skandhan, 1992; Huang et al., 2000; Massanyi et al., 2004; Yuyan et al., 2007). Our previous *in vitro* studies evaluated the negative effects of a wide range of concentrations of Cu as a risk factor of environment on the motility of spermatozoa and subsequent pointed out a cytotoxic effect of Cu on the mitochondrial complex (Knazicka et al., 2012a,b).

Since, that Cu plays an essential role in spermatogenesis and fertility; this study was carried out to determine relationship of seminal plasma Cu concentration and spermatozoa motility.

## **MATERIAL AND METHODS**

### **Biological material**

Bovine semen samples were obtained from 13 adult breeding bulls (Slovak Biological Services, Nitra, Slovak Republic). The samples had to accomplish the basic quality criteria given for the corresponding breed. The semen was obtained on a regular collection schedule using an artificial vagina. After collecting the samples, they were stored in the laboratory at room temperature (22-25 °C) and basic measurements were performed – volume (mL), pH, concentration ( $\times 10^9$ /mL) and osmolarity (mOsmol/kg) (Table 1). Each sample was diluted in physiological saline solution (PS) (sodium chloride 0.9% w/v, Bieffe Medital, Grosotto, Italia; pH – 5,5; osmolarity – 301 mOsmol/kg), using a dilution ratio of 1:40, depending on the original spermatozoa concentration.

### **Spermatozoa motility analysis**

The spermatozoa motility was carried out using the Computer Assisted Semen Analysis (CASA) system – SpermVision™ program (MiniTüb, Tiefenbach, Germany) with the Olympus BX 51 phase contrast microscope (Olympus, Tokyo, Japan) equipped with heating plate (37°C). Each sample was placed into the Makler Counting Chamber (depth 10  $\mu$ m, Sefi-Medical Instruments, Haifa, Israel) and the following parameters were evaluated: MOT - percentage of motile spermatozoa (%; motility > 5  $\mu$ m/s); PROG - percentage of progressive motile spermatozoa (%; motility > 20  $\mu$ m/s); DAP - distance average path ( $\mu$ m); DCL - distance curved line ( $\mu$ m); DSL - distance straight line ( $\mu$ m); VAP - velocity average path ( $\mu$ m/s); VCL - velocity curved line ( $\mu$ m/s); VSL - velocity straight line ( $\mu$ m/s); STR – straightness (VSL:VAP); LIN – linearity (VSL:VCL); WOB – wobble (VAP:VCL); ALH - amplitude of lateral head displacement ( $\mu$ m) and BCF – beat cross-frequency ( $H_z$ ).

### **Analysis of seminal plasma Cu concentration**

Subsequently, after measurements the samples were centrifuged (10 min, 9500 rpm, 4 °C) to obtain the cell sediment and seminal plasma fraction (supernatant). The analysis of Cu in the seminal plasma was performed BioLaTest (PLIVA-Lachema, Brno, Czech Republic) commercial kit. The measurement was based on a colorimetric reaction between Cu(I) ions and bathocuproine (BCP) forming a stable orange coloured complex, which was easy to detect photometrically at 480 nm (Genesys 10 spectrophotometer, ThermoFisher Scientific Inc., Madison, USA). Concentrations were expressed as µM/L.

### **Statistical analysis**

Statistical analysis of the results was carried out using the statistical program GraphPad Prism 3.02 (GraphPad Software Incorporated, San Diego, California, USA). Descriptive statistical characteristics (arithmetic mean, minimum, maximum, standard deviation and coefficient of variation) were evaluated. Pearson's correlation coefficient (two tailed) test was used to examine correlations between all the analyzed parameters of the semen. The level of significance was set at <sup>A</sup> ( $P<0.001$ ); <sup>B</sup> ( $P<0.01$ ); <sup>C</sup> ( $P<0.05$ ).

## **RESULTS AND DISCUSSION**

Semen volume, pH, concentration, viability and motility of spermatozoa as well as composition of the seminal plasma are common parameters to assess spermatozoa quality (Alavi and Cosson, 2006). These factors are directly related to the fertilization success (Bozkut et al., 2009).

The results of basic semen parameters showed that all observed characteristics were at physiological rates (Table 1). The semen has a very high buffering capacity, much higher than that of most other fluids in the body (Meacham, 2002). The bovine semen maintains a slightly acidic pH (Gamcik, 1992), which was in accordance with our results (pH of 6.57). The semen is notable also for its high osmolarity, which is substantially higher than that of blood plasma. The osmolarity of semen depends greatly on the concentration of sugars and other organics concentrations as well as ionic salt concentrations (Mandal and Bhattacharyya, 1987; Owen and Katz, 2005). In our experiment semen has a target

osmolarity of 297.20 mOsmol/kg. Some researchers have noted that osmolarity increases measurably with semen aging (Velazquez et al., 1977).

**Table 1** The basic parameters of analyzed bull semen samples (n=13).

PARAMETERS	x±S.D.
pH	6.57±0.18
Spermatozoa concentration (x10 <sup>9</sup> /mL)	3.42±0.96
Semen volume (mL)	6.25±2.45
Osmolarity (mOsmol/kg)	297.20±3.44
Motility (MOT; %)	92.46±3.99
Progressive motility (PROG; %)	90.23±4.02
Seminal plasma copper concentration (µM/L)	4.28±1.47

Legend: x – arithmetic mean; S.D. – standard deviation

The seminal plasma is a reliable biological marker for evaluating vitality, sperm metabolism, motility and others relevant semen parameters (Maxwell et al., 1996; Asadpour, 2012). Results of present study showed that the seminal plasma Cu concentration was in the range 2.14-6.89 µM/L with an average value of 4.28±1.47 µM/L. Eidi et al. (2010) examined seminal plasma levels of Cu and its relationship with human semen parameters. Their study demonstrated significant negative correlation between seminal plasma Cu concentration and pH ( $r_p = -0.173$ ;  $P < 0.05$ ) as well as sperm concentration ( $r_p = -0.114$ ;  $P < 0.05$ ). Subsequently, they confirmed that high concentration of Cu is related to lowering pH of seminal plasma, acidic pH, with changing condition of seminal plasma due to decrease motile or alive percent of spermatozoa. The excess Cu in seminal plasma is detrimental for male reproductive capacity by reducing spermatozoa count, motility, vitality and morphology.

The differences in opinion concerning the Cu content in seminal plasma of different species of animals were detected. The mean total Cu value of the buffalo seminal plasma in the study of Eghbali et al. (2008) was recorded as 2.51±0.04 mg/kg wet weight. Comparing with other authors we found out that according to Massanyi et al., (2003a,b) the seminal plasma Cu concentration was significantly higher ( $P < 0.01$ ) in the rams (2.49±0.18 mg/kg), fox (2.16±0.53 mg/kg) than that in the bulls (1.64±0.21 mg/kg), boars (1.64±0.28 mg/kg) and stallions (0.86±0.10 mg/kg). The concentration of Cu in rabbit semen was assessed Lukac et al. (2009) on the level 20.10±4.09 mg/kg wet weight, while rabbit semen is characterized by very high Cu concentration. Machal et al. (2002) state that the mean Cu level in seminal plasma of bulls was 38.17 µM/L, which in comparison with our results is too high a concentration.

**Table 2** The average values of spermatozoa motility parameters of analyzed bovine semen samples.

PARAMETERS MOTILITY	x	minimum	maximum	S.D.	CV (%)
MOT (%)	92.46	82.22	99.39	3.99	4.32
PROG (%)	90.23	80.55	96.66	4.02	4.46
DAP (µm)	37.73	29.26	54.07	5.08	13.48
DCL (µm)	60.78	47.83	88.77	8.34	13.73
DSL (µm)	31.43	23.21	47.82	4.92	15.65
VAP (µm/s)	89.42	68.47	137.70	12.76	14.27
VCL (µm/s)	143.70	110.90	208.10	20.20	14.06
VSL (µm/s)	74.83	55.93	123.40	12.21	16.32
STR	0.83	0.74	0.91	0.04	4.47
LIN	0.52	0.40	0.66	0.06	11.07
WOB	0.62	0.49	0.74	0.05	8.67
ALH (µm)	4.82	2.99	6.14	0.69	14.30
BCF (Hz)	34.65	27.41	44.60	3.36	9.70

Legend: MOT – percentage of motile spermatozoa (%); PROG - percentage of progressive motile spermatozoa (%); DAP- distance average path (µm); DCL – distance curved line (µm); DSL – distance straight line (µm); VAP- velocity average path (µm/s); VCL – velocity curved line (µm/s); VSL – velocity straight line (µm/s); STR – straightness (VSL:VAP); LIN – linearity (VSL:VCL); WOB – wobble (VAP:VCL); ALH - amplitude of lateral head displacement (µm) and BCF – beat cross-frequency (Hz).

x – arithmetic mean, S.D. – standard deviation, CV (%) – coefficient of variation

The mean value for the percentage of motile spermatozoa (quantity of movement) was  $92.46 \pm 3.99\%$ . The CASA analysis showed  $90.23 \pm 4.02\%$  of progressive motile spermatozoa (quality of movement). The other motility parameters with statistical differences are shown in the Table 2. **Eghbali et al. (2008)** recorded, that spermatozoa motility was  $92.24 \pm 0.51\%$  in excellent group,  $81.66 \pm 0.62\%$  in good group and moderate group  $71.66 \pm 1.05\%$ , which were significantly different.

The high concentration of Cu in seminal plasma is correlated with reduced spermatozoa motility (**Rebrelo et al., 1996; Eidi et al., 2010**). In our case, the correlation analysis revealed a strong negative correlation between percentage of motile spermatozoa and seminal plasma Cu concentration ( $r_p = -0.781$ ;  $P < 0.01$ ) as well as between progressive of motile spermatozoa and Cu content in the seminal plasma ( $r_p = -0.726$ ;  $P < 0.01$ ), which is in agreement with the report of **Eidi et al. (2010)** and **Akinloye et al. (2011)**. The findings of other authors are however controversial in the comparison with our results. **Eghbali et al. (2008)** demonstrated a positive correlation between seminal plasma Cu concentration and spermatozoa motility with viability. **Machal et al. (2002)** reported a statistically significant ( $P < 0.05$ ) positive coefficients of correlation between the Cu concentration in seminal plasma and spermatozoa motility ( $r_p = 0.330$ ) and the total number of sperm cells with progressive

motility ( $r_p=0.280$ ). These their results correspond with the studies of **Dhami et al. (1994)** and **Leonhard-Marek (2000)**. In a similar study, **Jockenhövel et al. (1990)** showed significant correlation between seminal plasma Cu concentrations and spermatozoa count, motility and normal morphology. The toxic effects of Cu on the seminal plasma are manifested in the decrease of motile spermatozoa percentage and in the increase of malformed spermatozoa (**Gamcik et al., 1990; Vrzgulova et al., 1993; Massanyi et al., 2005**).

**Table 3** The Pearson's coefficient of correlations ( $r_p$ -values) for relationship between seminal plasma Cu concentration and selected spermatozoa motility parameters.

	Cu	MOT	PROG
Cu	1		
MOT	-0.781 <sup>B</sup>	1	
PROG	-0.726 <sup>B</sup>	0.962 <sup>A</sup>	1

Legend: The correlation analysis was based on the value of the correlation coefficient:  $\pm 0.111$  to  $\pm 0.333$ : *low correlation*;  $\pm 0.334$  to  $\pm 0.666$ : *moderate correlation*;  $\pm 0.667$  to  $\pm 0.999$ : *high correlation*.

MOT – percentage of motile spermatozoa (%); PROG - percentage of progressive motile spermatozoa (%); <sup>A</sup> $P < 0.001$ ; <sup>B</sup> $P < 0.01$ ; <sup>C</sup> $P < 0.05$

## CONCLUSION

The data obtained from this study clearly indicated that concentration of copper in seminal plasma negatively affects the spermatozoa motility parameters and subsequently might cause reproductive alteration in male sexual functions. Therefore, it is important to systematically evaluate its presence in the seminal plasma.

**Acknowledgments:** The authors are grateful to Slovak Biological Services (Lužianky, Slovak Republic) for biological materials. This work was supported by the Tatra Bank Foundation 2012/2013 (Slovak Republic), by the Scientific Agency of the Slovak Republic VEGA No. 1/0532/11 and by KEGA Cultural and Educational Grant Agency no. 013SPU-4/2012.

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