NUTRITIONAL AND TECHNOLOGICAL PROPERTIES OF SELECTED KINDS OF COFFEE

Eva Ivanisová1*, Martina Czakóová1, Miroslava Kačaniová2,3, Eva Tvrdá4

Address(es):
1Department of Technology and Quality of Plant Products, Faculty of Biotechnology and Food Sciences, Slovak University of Agriculture in Nitra, Tr. A. Hlinku 2, SK-949 76.
2Department of Food Sciences, Viticulture and Enology, Faculty of Horticulture and Landscape Engineering, Slovak University of Agriculture in Nitra, Tr. A. Hlinku 2, SK-949 76.
3Department of Bioenergetics and Food Science, Faculty of Biology and Agriculture, University of Rzeszów, Zelewrowicza, , 4, PL-35-601.
4Department of Animal Physiology, Faculty of Biotechnology and Food Sciences, Slovak University of Agriculture in Nitra, Tr. A. Hlinku 2, SK-949 76.

*Corresponding author: eva.ivanisova@uniag.sk

ABSTRACT

The aim of the study was to evaluate the quality indicators (moisture content, fat, ash, polyphenols and antioxidant activity) of seven samples of green coffee. The coffee samples were subsequently roasted, using the cinnamon roasting style. In the prepared roasted coffee, the same measurements were made as in the green coffees, in order to compare the samples and point out the changes taking place during roasting. In addition, oxidative stability was determined after the roasting process. The moisture content of the green coffee ranged from 7.4 % to 9.08 %, the ash content ranging from 2.53 % to 2.97 %, the fat content from 2.19 % to 6.33 %. The DPPH antioxidant activity ranged from 45.36 to 55.81 mg TEACL1 (TEAC – Trolox equivalent antioxidant capacity) and total polyphenols from 511.44 to 618.11 mg GAE.l–1 (GAE – gallic acid equivalent). After the roasting process, the amount of dry matter (97.88 – 98.54 %) and fat (3.38 – 12.76 %) increased, the ash content did not change significantly. The most pronounced was the decrease in polyphenols, which is not surprising due to thermolabile substances. Overall, their values decreased of about 80 % compared to green coffee samples. Despite these losses, the samples showed a high antioxidant activity after the roasting process – 42.56 to 55.63 mg TEACL1, which can be explained by the fact that during the roasting the Maillard reaction takes place, whereby new bioactive substances with antioxidant effect arise, on activity it also participates in vitamin B1, which is produced by thermal breakdown of trigoneline. The oxidative stability evaluated by the Rancimat method ranged from 1.2 to 10 minutes, suggesting that roasted coffee is susceptible to oxidation, therefore it should be milled just before preparation and serving.

Keywords: Coffea sp., roasting, fat, oxidative stability, antioxidant activity, polyphenols

INTRODUCTION

Coffee is considered the most important alimentary raw material in the world and is produced extensively in about 60 tropical and subtropical countries (Cruz et al., 2017). Coffee is a main dietary source of polyphenol and phenolic acid due to its high polyphenol and phenolic acid content. These constituent of the coffee are correlated well with high antioxidant properties, weight loss, mood enhancing and increase alertness, effectiveness against hypertension and anticancer properties. Recently, the demand and consumption of green coffee bean has skyrocketed due to health properties contain in it (Zain et al., 2018). Green coffee beans are rich in the phenolic compounds exemplified by chlorogenic acid, caffeic acid, ferulic acid and p-coumaric acid. Coffee is the major source of chlorogenic acid in the human diet. On the basis of 10 g of coffee per cup of brew, a cup contains 15–325 mg of chlorogenic acids; daily intake of coffee drinkers is 0.5–1.0 g, whereas coffee abstainers typically ingest < 100 mg/day (Votavová et al., 2009). Bioactive compound composition in coffee beans and coffee brew are highly dependent on the processing steps, and roasting is the most crucial of these steps. High temperature and low water activity in the roasting process facilitate molecular degradation and the formation of new compounds (Herawati et al., 2019). However, roasting process introduced causes 8–10% chlorogenic acid degradation and transformation per each 1% loss of dry matter and 11% to 45% polyphenol degradation (Zain et al., 2018). The roasting process involves the application of heat to the beans ranging from 200 to 240 ºC for a period of time that depends on the desired final product. There are three main levels in coffee roasting, light, medium, and dark. The main difference in the process is the time that the coffee remains in the roaster and this process can affect the composition of the coffee indifferent ways (Leric and Nicoli, 1990). Roasting temperatures used, range from 160 to 240ºC for durations of 8 to 24 minutes. The cinnamon (bright) type of roasting coffee is characterised by bright cinnamon-coloured grains with a matt surface, obtained by short roasting and/or at lower temperatures. Such a shortly roasted coffee gives a mildly spicy brew (Przybysz et al., 2013).

The aim of this study was to determine technological and nutritional properties of seven coffees (green and after roasting process) originating from various regions in order to elucidate changes and differences between green and roasting coffee.

MATERIAL AND METHODS

Biological material and sample preparation

The seven coffee samples – Elite (90 % Arabica+10 % Robusta), Forte (50 % Arabica+50 % Robusta), Robusta India Kaapi Royal (100 % Robusta), Arabica Nicaragua Jutodog (100 % Arabica), Peru Jose Santos Campos (100 % Arabica), Tanzania Iyela PB (100 % Arabica), Kihangu United Uganda Rwenzori Mountains (100 % Arabica) were provided by the private supplier from Nitra (Slovakia). Green coffee (100 g) was fed to a micro-roaster coffee machine (Gene Café, CBR-101, Korea). The beans were roasted by using cinnamon roasting style – 235 ºC, 13 minutes. Roasted coffee bean was ground in a grinder machine (Krupps, GVX 2, Bur Grinder, Germany). Samples were ground to an average particle size (20 mesh), suitable for preparation in a paper filter. To prepare the extract, 0.1 g of sample was extracted with 20 ml 95 ºC distilled water, after 5 minute extraction filtered through filtration paper (Whatman no 1) and used for analysis (antioxidant activity, total polyphenol content).

Chemical

All the chemicals used were of analytical grade and were purchased from Sigma-Aldrich (St. Louis, MO, USA) and CentralChem (Slovakia).
Moisture content and ash content were determined according to the standard AACC method 08-01 (AACC 1996). Fat content was detected with Ancom XT15 Fat Extractor (USA) in line with manufacturer’s recommendation. One 1.5 g w.t. was weighted to special filter bag (XT4, Ancom, USA) and dried for 3 hours in an oven (WTB, Binder, Germany) at 105 °C to remove moisture prior to the extraction. Samples were placed in a desiccant pouch for 15 minutes and after re-weighted (W2) and extracted 60 minutes at 90 °C with petroleum ether. After process samples were removed and dried in an oven at 105 °C for 30 minutes, placed in desiccant pouch and re-weighted (W3). Fat content (%) was calculated by following formula: FC (%)=[(W2-W3)/W1]x100.

The oxidative stability was determined in 892 Rancimat apparatus from Mettler (Switzerland) according to ISO 6886:1997 utilizing a sample of 0.5±0.01 g. All samples were analyzed in triplicate, mg kg cemented at 120 °C under a constant air flow (20 L/h). The induction times were printed automatically by apparatus software with the accuracy of 0.005.

Antioxidant activity – DPPH method
Antioxidant activity of samples was measured using 2,2-diphenyl-1-picrylhydrazyl (DPPH) according to the procedures described by Sanchez-Moreno et al., (1998). An amount of 0.4 ml of extract was added to 3.6 ml of DPPH solution (0.025 g DPPH in 100 ml ethanol). Absorbance of the reaction mixture was determined using the spectrophotometer Jenway (6405 UV/Vis, England) at 515 nm. Trolox (6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid) (10-100 mg·L-1) Trolox equivalents were used as the standard and the results were expressed in mg·L-1 Trolox equivalents.

Total polyphenol content
Total polyphenol content was measured in accordance to Singleton and Rossi, (1965) using Folin-Ciocalteu reagent. A 0.1 ml of sample was mixed with 0.1 ml of the Folin-Ciocalteu reagent, 1 ml of 20% (w/v) sodium carbonate and 8.8 ml of distilled water, and left in darkness for 30 min. The absorbance at 700 nm was measured using the spectrophotometer Jenway (6405 UV/Vis, England). Gallic acid (25-300 mg·L-1; R2=0.998) was used as a standard and the results were expressed in mg·L-1 of gallic acid equivalent.

Statistical analysis
All experiments were carried out in triplicate and the results reported are the results of those replicate determinations with standard deviations. The experimental data were subjected to analysis of variance (Duncan’s test), at the confidence level of 0.05, by the use of software SAS (2009).

RESULTS AND DISCUSSION

Technological and nutritional evaluation of green coffee samples
Moisture content (Tab. 1) of analyzed green coffee samples ranged from 7.4 to 9.08%. Due to low level of moisture in coffee beans belong to stable crops, but this parameter must be monitored regularly. The bad storage condition a manipulation (especially higher moisture content) can cause growing of microscopic filamentous fungi in green beans. Viegas et al., (2017) in their study evaluated twenty-eight samples of green coffee (Arabica and Robusta) from different countries (Honduras, India, Costa Rica, Nicaragua) and found that the dominant contaminant in these samples was Aspergillus species (39 % Aspergillus niger and 29 % Aspergillus circundati). This microscopic filamentous fungus can produces mycotoxines, which are stable during roasting process, and can be poisonous for coffee consumer. From this reason is very important during the coffee processing to dry green coffee to required moisture content 10-12.5% and next storage in dry condition and lower air humidity.

Ash content in green coffee samples (Tab. 1) ranged from 2.53 to 2.97 %. In this work was determined total content of ash, but according to Farah, (2012) the dominant mineral element found in coffee is potassium. Its amount ranges from 1 to 2 g·100 g-1 which represents 40 % of the total mineral content. Phosphorus is nutritionally important and is attributed to the ability to lower high blood pressure and also relieves muscle cramps (Hoffmann, 2014). In addition to potassium, phosphorus is also present in coffee beans in amount approximately 4%. The coffee beans also contain microelements presented in small amounts – iron, copper, zinc but also magnesium, the amount of which is greatly influenced by the environment in which coffee plants are grown. Coffe plants grown on volcanic soil, for example, are characterized by a higher content of these elements (especially iron, copper, magnesium).

Fat content (Tab. 1) in analyzed green coffee samples ranged from 2.19 to 6.33 %. While the difference in moisture and ash content was minimal, the difference in the determination of fat was significant. For example in sample Robusta India Kaapi Royal was determined the amount of fat 2.19 % which was also the lowest value; in sample Peru Jose Santos Campos the amount of fat was 6.33 %. The average amount of fat was 4.65%.

Antioxidant activity and total polyphenol content of green coffee samples
Antioxidant activity determined by DPPH in analyzed samples of green coffee (Tab. 1) ranged from 45.36 to 55.81 mg TEAC·1. Sentkowski et al., (2016) similarly like in this study analyzed antioxidant activity (using DPPH method) of green coffee samples from different geographical origin. The samples of Arabica from Vietnam, Indonesia, Laos and Robusta from Vietnam, India, Indonesia, Laos and Uganda. The highest activity was determined in sample of Robusta from Laos 100 mg TEAC·1 and in the sample of Robusta from India 94.8 mg TEAC·1. In samples of Arabica was detected lower antioxidant activity with compare to Robusta, with the highest values in sample Arabica from Vietnam 94.8 mg TEAC·1 and the sample of Arabica from Laos 94.8 mg TEAC·1. Our results are comparable with results obtained in this study. Antioxidant activity of coffee is very strongly influenced by agro ecological condition, soil and geographical condition (Sentkowski et al., 2016; Zain et al., 2018; Herawati et al., 2019).

Coffee in our diet as a favorite soft drink plays an important role as it provides us with a significant amount of bioactive substances and thus contributes to the daily intake of antioxidants (Neduchalová, 2013). The total polyphenol content (Tab. 1) in analyzed green coffee samples ranged from 511.44 mg GAE·1 to 618.11 mg GAE·1. The highest value was detected in sample of Arabica Peru Jose Santos Campos, while the lowest value was determined in Arabica Tanzania Ivela PB. Similarly like antioxidant activity, amount of polyphenols in coffee beans is very strongly influenced by agro ecological condition, farming, soil and geographical condition (Sentkowski et al., 2016; Zain et al., 2018; Herawati et al., 2019).

Technological and nutritional evaluation of roasted coffee samples
Moisture content (Tab. 2) in analyzed roasted coffee samples (after roasting process – cinnamon roast) ranged from 1.46 to 2.12 %. After roasting process moisture content decreased, because due to high temperature the water content decreased, the volume of grains and friability increased – green coffee beans are hard, and it is almost impossible to chew them, but after the roasting process they are brittle and can be easily chewed. Thanking to the low moisture content the roasted coffee beans are stable product, but it should be properly stored, especially in a dry, well-closed packaging to avoid their contamination.

Fat content (Tab. 2) in analyzed roasted coffee samples (after roasting process – cinnamon roast) ranged from 2.95 to 3.38 %. The concentration of the individual elements increased explaining the moisture content and dry matter content was changed. Van Cuong et al., (2014) determined the influence of roasting process (temperature 210 °C – 20 minutes, 220 °C – 18 minutes, 230°C – 14 minutes, 240°C – 12 minutes and 250 °C – 8 minutes) to concentration of mineral elements – potassium, calcium and magnesium with green coffee beans. The concentration of the individual elements increased with compared to green coffee and the highest concentration was reached at a roasting temperature of 250 °C. Potassium (K), which was highest in green coffee, increased from 13.42 – 14.47 mg·g-1 to 13.84 – 15.09 mg·g-1. The concentration of calcium (Ca) also increased from 0.77 to 1.26 mg·g-1. The actual composition and content of the individual mineral elements is different and their content mainly affects the profile of the soil in which the coffee plants were grown (Sarapatka, 2014).

Fat content (Tab. 2) in analyzed roasted coffee samples (after roasting process – cinnamon roast) ranged from 3.58 to 12.76 %. The fat content of Arabica and Robusta coffee beans is different and ranges between 3-15%. Arabica contains generally higher amount of fat which was confirmed also in our study, whereas amount of fat was the lowest in sample of Robusta India Kaapi Royal. The endosperm of coffee bean contains the largest proportion of fat, with only a small portion of the fat situated in the surface layers of the beans (Speer, 2006). In terms of coffee fat composition, it is predominantly a mixture of triacylglycerols, di- and fatty acids. From the fatty acids, linoleic acid and palmitic acid are predominant, while from diacetylenes are predominant cafestol and kahweol. These have been increasingly studied in recent years because of physiological effects on the human body. A recent study of the effects of coffee di- and triacetylenes on the migration of prostate cancer cells confirmed their beneficial effect. Kahweol and cafestol synergistically inhibit the proliferation and migration of prostate cancer cells, therefore these coffee compounds may represent a potential solution to treat prostate cancer problems (Hiroaki et al., 2018). The difference between Arabica and Robusta is mainly in the content of 16-O-methylcafeestol, whose content has been found to be higher in Arabica only. 16-O-methylcafeestol is a reliable indicator for the detection of Robusta in coffee mixtures (Speer, 2006).

Antioxidant activity (Tab. 2) in analyzed roasted coffee samples (after roasting process – cinnamon roast) ranged from 42.56 to 55.63 mg TEAC·1. From results can be possible to see, that after roasting process (high temperature) is still content of polyphenolic compounds in beans, which are responsible for antioxidant activity of coffee. A major contributor to the antioxidant activity was identified as 15-methylpyridinium. The levels of 15-methylpyridinium in roasted and ground
Coffee is positively correlated to the degree of roasting (Votavová et al., 2009). Methylpyridinium is not present in raw coffee beans but it is formed during the roasting process from its chemical precursor, trigonelline, which is common in raw coffee beans. Trigonelline is the second most abundant alkaloid in green coffee and reaches levels in Arabica and Robusta from 7.9 to 10.6 g kg⁻¹ and from 6.6 to 6.8 g kg⁻¹, respectively (Stennert and Maier, 1994). Our results are comparable with the results of Neduchlova, (2013) which detected antioxidant activity by DPPH method in sample of Arabica Nicaragua Jinotega with value 50.52 mg TEAC and in sample Robusta India Kaapi Royal – 41.01 mg TEAC. The total polyphenol content (Tab. 2) in analyzed roasted coffee samples (after roasting process – cinnamon roast) ranged from 93.67 to 124.4 mg GAE.¹ Total polyphenols in roasted coffee significant decreased with compare to green coffee and this decrease was in level of ~80 % in all analyzed samples. Roasting evokes several changes in the constituents of coffee beans through modification or degradation. During roasting, high temperatures result in polyphenol degradation. Chlorogenic, malic and citric acids levels decreased, whilst quinic acid increases due to chlorogenic acid degradation. Thermal degradation of chlorogenic acids gives rise to phenolic compounds, such as chlorogenic acid lactones, which increase the bitter taste of coffee brews (Dybkowska et al., 2017). During roasting process chlorogenic acid decreased approximately from value 7 % in green coffee to value 4.5 % to roasted coffee. The content of chlorogenic acid varies from coffee to coffee, and the plant produces acid based on a number of stimuli, such as changes in climatic conditions or pest infestations. For this reason, the amount of chlorogenic acid in Robusta is higher because this coffee plant is better able to withstand weather fluctuations and is more resistant to external influences and pest infestations. Roasted Arabica coffee contains approximately 35-100 mg 100 ml of chlorogenic acid and Robusta approximately 55-175 mg 100 mg 100 ml (Stalmach, 2012).

Oxidation stability (Tab. 2) of analyzed roasted coffee ranged from 1.2 to 10 minutes and results confirmed that coffee due to higher fat content is very sensitive for oxidation, especially after milling process. Therefore, mainly milled coffee should not be stored for a long time and if so in the dark, in the cold, well packaged especially in a vacuum package. However, skilled barists know that the best coffee is made from freshly ground beans. Grinding into the long-term storage of ground coffee is manifested by formation of unpleasant sensory compounds (oxidized, bitterness, “fish” odors).

<table>
<thead>
<tr>
<th>Table 1</th>
<th>The results of moisture, ash, fat, total polyphenol content and antioxidant activity of green coffee beans</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sample</td>
<td>Moisture content [ %]</td>
</tr>
<tr>
<td>Elite (90 % Arabica +10 % Robusta)</td>
<td>9.01 ±0.02a</td>
</tr>
<tr>
<td>Forte (50 % Arabica + 50 % Robusta)</td>
<td>8.88 ±0.03a</td>
</tr>
<tr>
<td>100 % Arabica India Kaapi Royal</td>
<td>9.68 ±0.02a</td>
</tr>
<tr>
<td>100 % Arabica Nicaragua Jinotega</td>
<td>8.76 ±0.06bc</td>
</tr>
<tr>
<td>100 % Arabica Peru Jose Santos Campos</td>
<td>8.39 ±0.06ab</td>
</tr>
<tr>
<td>100 % Arabica Tanzania Ivela PB</td>
<td>8.27 ±0.01a</td>
</tr>
<tr>
<td>100 % Arabica Kihungu United Uganda Rwenzi Mountains</td>
<td>7.4 ±0.02ab</td>
</tr>
</tbody>
</table>

**TEAC** – Trolox equivalent antioxidant capacity; **GAE** – gallic acid equivalent; ± standard deviation; different letters in column denote mean values that statistically differ one from another.

<table>
<thead>
<tr>
<th>Table 2</th>
<th>The results of moisture, ash, fat, total polyphenol content, antioxidant activity and oxygen stability of analyzed roasted coffee samples</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sample</td>
<td>Moisture content [ %]</td>
</tr>
<tr>
<td>Elite (90 % Arabica +10 % Robusta)</td>
<td>2.12 ±0.12a</td>
</tr>
<tr>
<td>Forte (50 % Arabica + 50 % Robusta)</td>
<td>1.84 ±0.02a</td>
</tr>
<tr>
<td>100 % Robusta India Kaapi Royal</td>
<td>1.87 ±0.04ab</td>
</tr>
<tr>
<td>100 % Arabica Nicaragua Jinotega</td>
<td>1.81 ±0.15bc</td>
</tr>
<tr>
<td>100 % Arabica Peru Jose Santos Campos</td>
<td>2.04 ±0.11ab</td>
</tr>
<tr>
<td>100 % Arabica Tanzania Ivela PB</td>
<td>1.71 ±0.08a</td>
</tr>
<tr>
<td>100 % Arabica Kihungu United Uganda Rwenzi Mountains</td>
<td>1.46 ±0.07d</td>
</tr>
</tbody>
</table>

**TEAC** – Trolox equivalent antioxidant capacity; **GAE** – gallic acid equivalent; ± standard deviation; different letters in column denote mean values that statistically differ one from another.

**CONCLUSION**

This study showed that roasting process of coffee had influence for nutritional and technological parameters of coffee beans. During the roasting process decreased amount of moisture content and total polyphenol content, while amount of fat and mineral compounds increased. Antioxidant activity of analyzed samples was not changed significantly due to the components which is forming due to Mailard’s reaction and thanking to the degradation of trigonelline alkaloid. Results also showed, that roasted coffee due to higher amount of fat is very sensitive to oxidation, and so it is very important storage coffee with accordance to right storage conditions, especially ground coffee.

**Conflicts of interest:** All authors declare no conflicts of interest.

**Acknowledgments:** This work was co-funded by the KEQA project 009SPU-4/2018 (90 %) and project 05-GA SPU-16 (10 %).

**REFERENCES**


